

## **Phosphorylation- and Nucleotide-Binding- Induced Changes to the Stability and Hydrogen Exchange Patterns of JNK1 $\beta$ 1 Provide Insight into Its Mechanisms of Activation**

Gavin R. Owen<sup>1</sup>, Stoyan Stoychev<sup>2</sup>, Ikechukwu Achilonu<sup>1</sup> and Heini W. Dirr<sup>1</sup>

1 Protein Structure–Function Research Unit, School of Molecular and Cell Biology, University of the Witwatersrand, Johannesburg 2050, South Africa

2 Biosciences, Council for Scientific and Industrial Research, Pretoria 0001, South Africa

### **Abstract**

Many studies have characterized how changes to the stability and internal motions of a protein during activation can contribute to their catalytic function, even when structural changes cannot be observed. Here, unfolding studies and hydrogen-deuterium exchange (HX) mass spectrometry were used to investigate the changes to the stability and conformation/conformational dynamics of JNK1 $\beta$ 1 induced by phosphorylation. Equivalent studies were also employed to determine the effects of nucleotide binding on both inactive and active JNK1 $\beta$ 1 using the ATP analogue, 5'-adenylyl imidodiphosphate (AMP-PNP). JNK1 $\beta$ 1 phosphorylation alters HX in regions involved in catalysis and substrate binding, changes that can be ascribed to functional modifications in either structure and/or backbone flexibility. Increased HX in the hinge between the N- and C-terminal domains implied that it acquires enhanced flexibility upon phosphorylation that may be a prerequisite for interdomain closure. In combination with the finding that nucleotide binding destabilizes the kinase; the patterns of solvent protection by AMP-PNP were consistent with a novel mode of nucleotide binding to the C-terminal domain of a destabilized and open domain conformation of inactive JNK1 $\beta$ 1. Solvent protection by AMP-PNP of both N- and C-terminal domains in active JNK1 $\beta$ 1 revealed that the domains close around nucleotide upon phosphorylation, concomitantly stabilizing the kinase. This suggests that phosphorylation activates JNK1 $\beta$ 1 in part by increasing hinge flexibility to facilitate interdomain closure and the creation of a functional active site. By uncovering the complex interplay that occurs between nucleotide binding and phosphorylation, we present new insight into the unique mechanisms by which JNK1 $\beta$ 1 is regulated.