

1 **Antileishmanial activity of 12-methoxycarnosic acid from**

2 ***Salvia repens* Burch. ex Benth. (Lamiaceae)**

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21 **Abstract**

22 In South Africa, *Salvia repens* is used traditionally to treat sores, stomach ache and diarrhoea.
23 The high performance liquid chromatography (HPLC)-based activity profiling of *S. repens*
24 whole plant extract showed an active abietane diterpene which was identified as 12-
25 methoxycarnosic acid (**1**) which showed antiprotozoal activity against axenically grown *L.*
26 *donovani* amastigotes with IC₅₀ of 0.75 µM with marginal cytotoxicity against the L6-cells (IC₅₀,
27 17.3 µM).

28

29 **Keywords:** Antileishmanial activity; *Salvia repens*; *Leishmania donovani*, 12-methoxycarnosic
30 acid, abietane diterpene, HPLC profiling

31

32 **Abbreviations:** VL = Visceral leishmaniasis; HIV = Human Immunodeficiency virus; SANBI =
33 South African National Biodiversity Institute; HPLC = high performance liquid chromatography;
34 DMSO =dimethyl sulfoxide; PBS = phosphate buffer solution; FBS = fetal bovine serum; PDA =
35 Photo Diode Array; NMR = Nuclear Magnetic Resonances; MS = Mass Spectrometry; SI =
36 selectivity index; DHT = dihydrotestosterone; AR = androgen receptor; SM = simple
37 monophasic.

38 1. Introduction

39 Visceral leishmaniasis (VL), also known as “Kala azar” is caused by the intracellular protozoan
40 *Leishmania donovani* and is transmitted by the bite of infected phlebotomine sandflies. It has
41 been estimated that the annual global burden of VL is 500 000 new cases with more than 59 000
42 associated deaths (Den Boer et al., 2011). The control of VL relies entirely on chemotherapy
43 due to lack of antileishmanial vaccines. In poor countries, however, the use of leishmanicidal
44 drugs is hampered by high costs, toxicity and the emergence of resistance (Sundar et al., 2007).
45 In addition, the emergence of the Human Immunodeficiency virus/Acquired Immunodeficiency
46 disease (HIV/AIDS) has severely compromised the control of VL (Alvar et al., 2008).
47 Therefore, there is an urgent need for the development of new leishmanial therapeutics.

48 The genus *Salvia* (sage) is the largest in the Lamiaceae family with over 900 species worldwide
49 (Tan et al., 2002). Approximately 30 *Salvia* species occur in southern Africa of which 26 are
50 found in South Africa especially in the Cape region (Codd, 1985). *Salvia* species are used in
51 folk medicine for the treatment of asthma, eczema, psoriasis and tuberculosis (Salimpour et al.,
52 2011). In South Africa, *Salvia* species are used against fever, headache and digestive disorders
53 (Amabeoku et al., 2001), and to treat sores (Auge et al., 2003). A decoction of the roots is also
54 used in South Africa for the treatment of stomach ache and diarrhoea (Auge et al., 2003).
55 Several kinds of tanshinone-type diterpenoids (Ślusarczyk et al., 2011) abietane and icetexane
56 diterpenoids and triterpenoic acids (Nieto et al., 2000; Tan et al., 2002; Kabouche et al., 2007),
57 phenolic acids, phenolic glycosides, flavonoids and anthocyanins (Walker and Sytsma, 2007)
58 have been reported from *Salvia* species. These constituents have shown various biological
59 properties including antiviral (Tada et al., 1994), antiprotozoal (Sairafianpour et al., 2001;

60 Ślusarczyk et al., 2011; Farimani et al., 2012) antimalarial (Achenbach et al., 1992),
61 antileishmanial (Tan et al., 2002) activities.

62 In our previous study, the dichloromethane/methanol (1:1) extract of the whole plant of *Salvia*
63 *repens* showed promising antiprotozoal activity with a IC_{50} value of 5.4 $\mu\text{g/ml}$ against axenically
64 grown *Leishmania donovani* amastigotes (MHOM/ET/67/L82) (Mokoka et al., 2011). In this
65 paper we report the isolation and identification of the active component and its antileishmanial
66 activity.

67

68 **2. Materials and methods**

69 **2.1. Plant collection**

70 *Salvia repens* whole plant material was collected from the Eastern Cape Province near Aliwal-
71 North on January 2002. The identification of plant material was done at the South African
72 National Biodiversity Institute (SANBI, Pretoria) where the voucher specimen (BP00998) was
73 deposited. The wet plant material was placed in an oven at 40°C for three days to dry, then
74 milled to a fine powder and kept at room temperature in the dark before extraction.

75

76 **2.2. Extract preparation**

77 Finely ground plant material of *S. repens* (100 g) was extracted with (2 × 1 L) of a mixture of
78 dichloromethane and methanol (1:1, v/v) overnight while occasionally aggitating. The mixture
79 was filtered and concentrated using a rotavapour at 40 °C and left under a stream of cold air to
80 dry.

81

82 **2.3. HPLC activity profiling**

83 The method described by Adams et al (2009) was used for the profiling of *S. repens* extract.
84 Briefly, 10 mg/ml sample in dimethyl sulfoxide (DMSO) was prepared, filtered through 0.45
85 µm sterile filters for HPLC fractionation. Gilson 215 liquid handler with Gilson 819 injection
86 module and 50 µl loop was used as the autosampler, connected to an HPLC system consisting of
87 a 1100 series low-pressure mixing pump with degasser module, column oven, and a 1100 series
88 PDA detector (Agilent, Waldbronn, Germany). The mobile system used was made up of water
89 with 0.1% formic acid (A) and acetonitrile with 0.1% formic acid (B). Separation conditions:
90 Sunfire RP-18 column (3.5 µm, 3 mm × 150 mm i.d.; Waters GmbH, Eschborn, Germany),
91 water (A) 90-0% in acetonitrile (B) over 30 min, 100% acetonitrile (B) for 5 min. The flow rate
92 was 0.5 ml/min, and the injection volume was 35 µl (350 µg extract in DMSO). Thirty-five one-
93 minute fractions were collected in 96 deep well plates during the run (Screenmates 96 well,
94 Matrix Technology, Hudson, USA) and dried in a GeneVac EZ-2 Plus evaporator (Genevac Ltd.,
95 Ipswich, UK). The samples were redissolved in 5 µl of DMSO and diluted with 95 µl phosphate
96 buffer solution (PBS) buffer (137 mM NaCl, 10 mM phosphate, 2.7 mM KCl, pH 7.4) for
97 microfraction antileishmanial screening.

98

99 **2.4. *In vitro* antileishmanial activity evaluation of the microfractions and isolated** 100 **compound**

101 The anti-leishmanial activity of the microfractions and isolated compound was evaluated by the
102 method described by Adams et al. (2009) at two different concentrations (5 µg/ml) and (0.8

103 $\mu\text{g/ml}$) against *L. donovani* amastigotes (MHOM/ET/67/L82) in 96-well microtiter plate. The
104 plate was read in a spectramax Gemini XS microplate fluorometer (Molecular Devices
105 Cooperation, Synnyvale, CA, USA) using an excitation wavelength of 536 nm and emission
106 wavelength of 588 nm. Fluorescence development was measured and expressed as a percentage
107 of the control. The IC_{50} value was determined from the sigmoidal inhibition curve and
108 miltefosine was used as a positive control.

109

110 **2.5. Cytotoxicity evaluations of 12-methoxycarnosic acid against L6-cells**

111 Assays were performed in 96-well microtiter plates, as previously discussed by Adams et al
112 (2009). The plates were incubated for 72 hours and thereafter inspected under an inverted
113 microscope to assure growth of the controls and sterile conditions. A volume of 10 μl of
114 resazurin was then added to each well and the plates incubated for a further 2 hours. The plates
115 were then evaluated as described above. The IC_{50} value was calculated from the sigmoidal
116 inhibition curve using SoftmaxPro software and podophylotoxin was used as a positive control.

117

118 **2.6. Isolation of the active compound**

119 The semi-preparative HPLC purification of the DCM/MeOH (1:1) crude extract of *Salvia repens*
120 whole plant was performed with an HPLC 1200 series consisting of a low-pressure mixing pump
121 with degasser module, a column oven and a PDA detector (Agilent, Waldbronn, Germany). A
122 100 mg/ml sample in DMSO was prepared and filtered through a 0.45 μm Millipore (Bedford,
123 MA, USA) membrane filter and several repetitions of 400 μl injections of the sample were made.

124 The separation was performed on a Waters Sunfire RP-18 column (10 × 150 mm i.d.; 10 µm;
125 Waters GmbH, Ireland). The mobile system used was similar to that used for the HPLC activity
126 profiling of the extract and made up of water (A) and acetonitrile (B). The elution gradient was
127 linear from A:B (90:10) to (0:100) in 30 min at a flow rate of 6.0 ml/min followed by washing
128 and returning to the initial elution conditions (90:10) over 5 min. The separation of the active
129 component was detected at 280 nm. The targeted fraction containing the compound of interest
130 was collected and dried in a Genevac evaporator overnight to afford compound **1**. The structure
131 of this compound was determined by ¹H and ¹³C nuclear magnetic resonances (NMR)
132 spectroscopy and mass spectrometry (MS) analysis as well as by comparison with the published
133 data and was identified as 12-methoxycarnosic acid (**1**) (Richheimer et al., 1996). Using 800 mg
134 of crude extract 10 mg of pure compound **1** was isolated. The purity was >97% as was shown by
135 HPLC.

136

137 **3. Results and discussion**

138 In our effort to identify potential antileishmanial compound/s from the DCM/MeOH (1:1) extract
139 of *Salvia repens* whole plant, an analytical HPLC-based activity profiling technique (Adams et
140 al., 2009) was used to fractionate the *S. repens* extract in a 96 deep-well microtiter plate. The
141 antileishmanial activity was in fraction 27 (figure **1**). The HPLC chromatogram at 280 nm
142 depicted in figure **1** made it possible to identify the active peak responsible for the observed
143 antileishmanial activity.

144 Using semipreparative HPLC a known compound was isolated and identified as 12-
145 methoxycarnosic acid (**1**) from its ¹H and ¹³C NMR spectra and by comparison of the NMR and

146 MS data with that published in the literature (Richheimer et al., 1996). The compound has been
147 isolated previously from *S. microphylla* (Aydoğmuş et al., 2006) and *Rosmarinus officinalis*
148 (Richheimer et al., 1996, Oluwatuyi et al., 2004).

149 The antileishmanial activity of 12-methoxycarnosic acid (**1**) was evaluated against axenic
150 *Leishmania donovani* amastigotes and showed an IC₅₀ value of 0.75 μM. Miltefosine (the
151 positive control) had an IC₅₀ of 0.19 μM. The selectivity index (SI), calculated as the ratio
152 between IC₅₀ value for cytotoxicity and IC₅₀ value for *L. donovani* amastigotes for compound (**1**)
153 was 23.2.

154 Previous reports have shown that **1** has very interesting biological activities. Murata and co-
155 authors (2012) reported that, **1** resulted in a 66.7% binding inhibition of 5α-dihydrotestosterone
156 (DHT) to the androgen receptor (AR) at 5 μM and the inhibition of 5α-reductase (IC₅₀, 61.7
157 μM), thereby playing a significant role in promoting hair growth. To the best of our knowledge
158 this is the first report on the antileishmanial activity of 12-methoxycarnosic acid (**1**).

159

160 **4. Conclusion**

161 The use of HPLC-based activity profiling made it possible to identify the active compound
162 responsible for antileishmanial activity. Compound (**1**), 12-methoxycarnosic acid isolated from
163 *Salvia repens* showed in *vitro* antileishmanial activity against axenic *L. donovani* amastigotes.
164 In addition, compound **1** has good selectivity. Further research into the structure of this
165 compound could result in the development of new analogues with much more improved activity
166 against this parasite.

167

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173

174 **Conflict of interest**

175 The authors declare that they have no conflict of interest.

176

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234 phylogenetic evidence for multiple origins of the lever. *Annals of Botany* 100, 375-391.

235 **Figure legend**

236 Figure 1. Antileishmanial activity trace of a DCM/MeOH (1:1) extract of the whole plant of
237 *Salvia repens* fractions and UV trace (280 nm) of the one minute fractions tested against
238 axenically grown *L. donovani* amastigotes at lower concentration (0.8 µg/ml) and the Chemical
239 structure of 12-methoxycarnosic acid (**1**)

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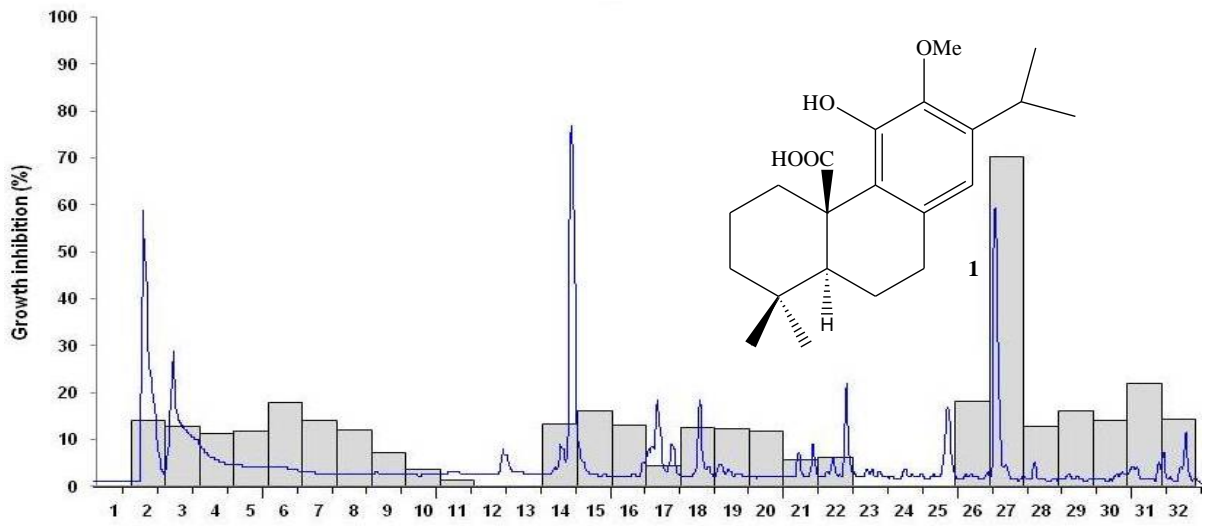
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248 **Figure 1**