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Impact of Prosopis invasion on a keystone tree species in the Kalahari Desert

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Abstract:	Several Prosopis species were introduced into South Africa in the last century. Here we determine the extent to which increased mortality of Acacia erioloba E.Mey, a keystone species in the Kalahari Desert of Southern Africa, can be attributed to competition for water with Prosopis. We do this through a determination of canopy vitality, plant water stress and plant water source. We use a visual estimate to determine plant vitality. Plant water stress we determine through a combination of stable carbon isotope ratios and xylem pressure potentials. Plant water source we determine using stable hydrogen and oxygen isotope ratios. Our results show that Prosopis abundance increases in the riparian zone and that there is a good correlation between greater Prosopis abundance and Acacia erioloba mortality. We show that both species are reliant on the same water resource in the riparian zone but that Acacia erioloba is better adapted to using the deeper water away from the river. We conclude that the decline in Acacia erioloba vitality at the river is related to competition for water with Prosopis. Our study gives strong support for the eradication of Prosopis from rivers in arid parts of Southern Africa.
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Prof Jacobson completed his PhD on arid rivers in Southern Africa and is still actively working in the area at times, being well-versed with the tree species that this submitted paper discusses

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Impact of *Prosopis* invasion on a keystone tree species in the Kalahari Desert

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ABSTRACT

Several *Prosopis* species were introduced into South Africa in the last century. Here we determine the extent to which increased mortality of *Acacia erioloba* E.Mey, a keystone species in the Kalahari Desert of Southern Africa, can be attributed to competition for water with *Prosopis*. We do this through a determination of canopy vitality, plant water stress and plant water source. We use a visual estimate to determine plant vitality. Plant water stress we determine through a combination of stable carbon isotope ratios and xylem pressure potentials. Plant water source we determine using stable hydrogen and oxygen isotope ratios. Our results show that *Prosopis* abundance increases in the riparian zone and that there is a good correlation between greater *Prosopis* abundance and *Acacia erioloba* mortality. We show that both species are reliant on the same water resource in the riparian zone but that *Acacia erioloba* is better adapted to using the deeper water away from the river. We conclude that the decline in *Acacia erioloba* vitality at the river is related to competition for water with *Prosopis*. Our study gives strong support for the eradication of *Prosopis* from rivers in arid parts of Southern Africa.

KEY WORDS

Kalahari Desert, water access, alien invasive, keystone species, *Acacia erioloba*, *Prosopis*

28 INTRODUCTION

29 Primarily introduced to provide shade or fodder in treeless, arid environments (Zimmerman
30 1991), several species of *Prosopis* (mesquite) have naturalized in desert regions throughout the world
31 (Burkhart 1976; Nilsen et al. 1991). In its native range *Prosopis* has become extremely invasive and is
32 now the dominant woody species on millions of hectares of semi-arid grassland (Brown and Archer 1989,
33 van Auken 2000). However, it is the effects of the various *Prosopis* species on natural vegetation in
34 Australia, the Indian subcontinent and 25 African countries (Mwangi and Swallow 2005) that has led the
35 IUCN to rank the genus among the world's worst invasive species (Mwangi and Swallow 2005;
36 Bromilow 2010). Around the turn of the 19th Century, four *Prosopis* species from central America were
37 introduced to Namibia and the north-western parts of South Africa (Harding 1987). Since then two
38 species, *Prosopis glandulosa* var *torreyana* (Benson) Johnson and *Prosopis velutina* Wooten have
39 densely and rapidly invaded riparian corridors and areas with shallow groundwater (Poynton 1990;
40 Zachariades et al. 2011; Wise et al. 2012). These species have also hybridised and may form impenetrable
41 thickets, making invaded farmland unproductive for crop production and livestock farming. *Prosopis* is
42 now spread over more than 1.45 million hectares in the Northern Cape Province of South Africa alone
43 (Wise et al. 2012).

44 *Acacia erioloba* E.Mey. is one of the most important woody species of the desert regions of
45 southern Africa (Coates Palgrave and Coates Palgrave 2002). Considered to be a keystone species in the
46 Kalahari *A. erioloba* provides shade, shelter, nesting sites and lookout posts for birds and other animals
47 (Milton and Dean 1995). There are, however, growing concerns that there is an increase in mature *A.*
48 *erioloba* mortality in the south western Kalahari. The proposed reasons for these concerns include an
49 increase in fire frequency (Seymour 2008), increasing groundwater abstraction (Powell, 2005) and
50 competition for resources from alien invasives such as *Prosopis* (Robertson and Woodborne 2002;
51 Zachariades et al. 2011).

52 In arid savannas such as the southern Kalahari, where mean annual precipitation is around 280
53 mm, rainfall is a strong constraint on the determinants of woody cover (Sankaran et al. 2005, 2008). As
54 an adaptation to survival in low rainfall environments, both *Prosopis* and *A. erioloba* have deep root
55 systems with records of *Prosopis juliflora* at 53 m (Canadell et al. 1996) and *A. erioloba* at 60 m
56 (Jennings 1974). These deep root systems allow these species to exploit deep aquifers in the sands on
57 which they grow. Recent stable isotope research has shown that *A. erioloba* may source groundwater as

58 deep as 70 metres below the surface (Obakeng 2007). The argument that *Prosopis* invasion is responsible
59 for *A. erioloba* mortality is based on the premise that both species are accessing deep groundwater with
60 *Prosopis* driving the water table down faster than *A. erioloba* may follow (Robertson and Woodborne
61 2002; Zachariades et al. 2011). Here we test this hypothesis by determining the relationship between
62 canopy vitality, plant water stress and plant water source for *A. erioloba* growing in conjunction with and
63 without *Prosopis*. We do this on a farm in the Northern Cape Province of South Africa that had been
64 partially cleared of *Prosopis*.

65

66 METHODS

67 *Site description*

68 The study was conducted on the farm Gannavlake (26 57.578 S- 21 50.234 E) in the southern
69 Kalahari, along the middle reaches of the ephemeral Kuruman River. While the river may flow in its
70 middle and lower reaches after unusually high rainfall (Meyer et al. 1985), the average groundwater depth
71 is 56 metres (Dept. of Water Affairs borehole records). The climate of the area is characterised by cold,
72 dry winters and hot, wet summers with average minimum and maximum temperatures of 13.2°C and
73 31.3°C respectively and an average annual rainfall of 280 millimetres (van Rooyen et al. 2001). The area
74 is largely covered with aeolian sand underlain by superficial silcretes and calcretes of the Cenozoic
75 Kalahari Group (Mucina and Rutherford 2006). The riverbed typically consists of finer silt soils than
76 those of the surrounding area, as water carries organic material, minerals and other alluvial components
77 into the lower lying riverbeds (van der Walt and le Riche 1999).

78 The landscape is shaped by parallel dunes about 3 to 8 metres high, which are vegetated by
79 *Stipagrostis amabilis* (Schweick.) De Winter, *Acacia haematoxylon* Willd. and *Acacia mellifera* (Vahl)
80 Benth (Mucina and Rutherford, 2006). The interdune plains are dominated by *Rhigozum trichotomum*
81 Burch. The river fringes are lined with riverine woodland, consisting primarily of *Acacia erioloba* and
82 other species such as *Acacia haematoxylon*, *Acacia mellifera*, *Boscia albitrunca* Burch., *Ziziphus*
83 *mucronata* Willd., *Rhigozum trichotomum* and *Prosopis* (van Rooyen et al. 2001).

84 *Woody plant density*

85 *Prosopis* was cleared in the Kuruman River, along half of the farm, by the national alien invasive plant
86 clearing programme 'Working for Water' (van Wilgen et al. 1998), while the other half remained un-

87 cleared. Before we established our plots we determined the density of all the trees in the cleared and un-
88 cleared (invaded) areas. For this, six transects were established, three in cleared and three in un-cleared
89 areas. Each transect was 5 metres wide and 300 metres long, moving diagonally away from the river.
90 Along each transect all mature trees were identified and counted.

91 Subsequent to the survey, four 20 m by 100 m plots were laid out, of which one was located along the
92 river and one 300 m away from the river in a upland invaded area. This setup was repeated in a cleared
93 area. The plots in the invaded area were densely overgrown with *Prosopis* while there were no mature
94 *Prosopis* in the cleared plots. In each plot we randomly selected six mature *A. erioloba* and six *Prosopis*.

95 ***Percentage canopy dieback***

96 In April 2007 two observers scored canopy dieback as a percentage of dead woody material
97 relative to the total canopy area on the study trees in each plot. Scores were made in 10% increments,
98 with 0% dieback a full healthy canopy and 100% dieback equating to a dead tree. The score average
99 between the two observers was recorded as the percentage canopy dieback for each tree.

100 ***Water source***

101 Sampling and measurements were carried out three times a year over two years, in the dry
102 season (June 2004 and July 2005), the early wet season (November 2004 and 2005) and the late wet
103 season (April 2004 and March 2005).

104 We use the stable isotopes of hydrogen and oxygen in water extracted from woody tissue to
105 show the water source for both *Prosopis* as well as *A. erioloba* (Busch et al. 1992; February et al.
106 2007a,b). The method is based on the assumption that water extracted from non-suberized wood will have
107 the same isotope ratio as the source water of the tree (White et al. 1985). We collected twig samples (c.
108 0.5 cm x 6 cm) from six randomly selected individual trees of each species in each plot. These twig
109 samples of non-suberized wood were collected into borosilicate tubes (Kimax–Kimble, Vineland, USA)
110 which were subsequently inserted onto a cryogenic vacuum extraction line to extract the xylem water for
111 isotope analysis. At the same time as the twig samples were collected, we also collected groundwater
112 samples from three different boreholes close to the study site. Rain water was collected during every
113 rainfall event and decanted into a sealed bottle after each event. During one wet season (April 2004) the
114 soil was sufficiently wet to auger 50 cm deep soil samples. To minimise evaporation this soil was put into

115 a polythene bag, inserted into a second bag, each of which was securely and individually sealed with
116 adhesive tape.

117 All water samples were analysed for $^2\text{H}/\text{H}$ ratios using a variation of the zinc closed tube
118 reduction method of Coleman et al. (1982), while $^{18}\text{O}/^{16}\text{O}$ ratios were obtained using the CO_2 equilibrium
119 method of Socki et al. (1992). Isotopic ratios of both $^2\text{H}/\text{H}$ and $^{18}\text{O}/^{16}\text{O}$ were then determined using a
120 Thermo Delta Plus XP Mass Spectrometer (Hamburg, Germany) at the University of Cape Town.

121 *Plant moisture stress*

122 *Leaf stable carbon isotope ratios*

123 Leaf carbon isotope ratios are correlated with leaf gas exchange. Carbon assimilation is
124 determined by stomatal aperture which in turn is determined by available water. As plants become more
125 water stressed, the stomatal aperture closes which results in less discrimination against the heavy ^{13}C
126 isotope, resulting in less negative $\delta^{13}\text{C}$ values (Ehleringer 1993). Using stable carbon isotope ratios we
127 establish the amount of water stress that both *Prosopis* and *A. erioloba* are under. In April 2007 a total of
128 twenty fully expanded, mature leaves were collected for stable carbon isotope analysis from each of our
129 six trees in each plot. Prior to mass spectrometry using a Thermo Delta Plus XP Mass Spectrometer
130 (Hamburg, Germany) the leaves were oven dried at 70°C for 24 hours and ground to a fine powder using
131 a Retsch MM200 ball mill (Retsch, Haan, Germany).

132

133 *Xylem Pressure Potentials*

134 Xylem pressure potentials are a determination of the tension that the water column of the plant is
135 under. The less water available to the plant the greater the tension on the water column (Scholander et al.
136 1965; Miller et al. 1984). We use predawn Xylem Pressure Potentials on our study trees to determine
137 plant moisture stress. We do this three times a year over two years, in the dry season (June 2004 and July
138 2005), the early wet season (November 2004 and 2005) and the late wet season (April 2004 and March
139 2005), using a Scholander type pressure chamber (PMS Instrument Company, Corvallis, Oregon, USA).

140

141 *Data analyses*

142 All statistical analyses for $\delta^{18}\text{O}$, $\delta^2\text{H}$, $\delta^{13}\text{C}$ isotopes and canopy dieback were conducted using
143 Statistica 8.0. We used Student T-tests and one-way ANOVA's with Tukey post hoc tests to detect any

144 significant differences ($p < 0.05$). A non-parametric Kruskal-Wallis test was used when assumptions of
 145 normality and heterogeneity of variance were not met. Repeated-measures ANOVA were used for
 146 predawn XPP in SPSS version 15.0, as described in ACITS (1997). Time was used as the within-subjects
 147 factor and tree groups as the between-subject factors. The multivariate hypothesis testing approach
 148 (Wilks' Lambda test) was used throughout.

149

150 RESULTS

151 *Woody plant density*

152 Our results for the belt transect show that *A. mellifera* and *Ziziphus mucronata* are absent from
 153 areas with high *Prosopis* density. In both the cleared and invaded transects *A. erioloba* density was
 154 highest immediately adjacent to the river with another increase in density approximately 300 metres from
 155 the river. *Prosopis* density was highest adjacent to the river, decreasing rapidly with increasing distance
 156 from the river. In the riparian zone there is a 50% increase in the number of *A. erioloba* trees where
 157 *Prosopis* had been cleared (Fig.1).

158 *Percentage canopy dieback*

159 There was a significant difference in the amount of dead material on *A. erioloba* between the
 160 different treatments. In the invaded river plot there was 50% more dieback than in the cleared river plot.
 161 As the number of *Prosopis* trees decline with distance from the river the amount of dead material on *A.*
 162 *erioloba* trees also declines ($F_{3,20} = 5.3$, $p = 0.008$) (Fig.2).

163 *Water source*

164 There is a linear relationship between $\delta^{18}\text{O}$ and $\delta^2\text{H}$ for fresh water samples described by the
 165 global meteoric water line (GMWL) with the equation $\delta^2\text{H} = 8\delta^{18}\text{O} + 10$ (GMWL) (Craig 1961).
 166 Evaporatively enriched water (shallow soil water) plots below the meteoric water line with a slope that is
 167 less than 8 and an intercept less than 10. We constructed a local meteoric water line (LMWL) from our
 168 rainfall data ($\delta^2\text{H} = 6.1 * \delta^{18}\text{O} + 2.6$ ‰). (Fig. 3). Values for groundwater isotope ratios are consistent
 169 over time and across the three sampling boreholes. The three soil core values (0.5 metre depth) show
 170 isotopic enrichment, relative to rainfall isotope ratios. These values plot below the LMWL (Fig. 3).

171 All isotope ratios for xylem water in the cleared river plot are located below the LMWL and
 172 intermediate between rainfall and groundwater (Fig. 3a). At the end of the wet season *A. erioloba* xylem

173 water isotope ratios differ significantly from groundwater (Kruskall-Wallis nonparametric $\delta^{18}\text{O}$; $p <$
 174 0.001). There are no significant differences between *A. erioloba* xylem water and ground water at the
 175 upland plot (Fig 3b).

176 Both *Prosopis* and *A. erioloba* xylem water isotope ratios in the invaded river plot (Fig. 3c)
 177 differ significantly from groundwater in the wet season (Kruskall-Wallis nonparametric $\delta^{18}\text{O}$; $p \leq 0.02$),
 178 but are not significantly different in the dry season.

179 In the invaded upland plot *A. erioloba* and *Prosopis* isotope ratios are similar to groundwater
 180 values (Fig. 3d). At the end of the wet season, however, values for *Prosopis* xylem water in this plot are
 181 significantly different from groundwater values (Kruskall-Wallis nonparametric $\delta^{18}\text{O}$; $p \leq 0.012$), but not
 182 significantly different from rain.

183 ***Plant moisture stress***

184 *Leaf stable carbon isotope ratios*

185 There were significant differences in $\delta^{13}\text{C}$ values between plots ($p < 0.001$). *Prosopis* $\delta^{13}\text{C}$
 186 values are significantly more enriched than values for *A. erioloba* (Fig. 4). The results for *A. erioloba*
 187 show significant differences in $\delta^{13}\text{C}$ values between cleared (-26.25‰) and invaded (-24.99‰) river plots
 188 ($p = 0.03$). There are no significant differences between cleared and invaded upland plots for *A. erioloba*.
 189 There are also no significant differences in $\delta^{13}\text{C}$ values for *Prosopis* invaded river and upland plots.

190 *Xylem pressure potentials*

191 Predawn XPP's for all upland plots are stable across seasons, relative to the river plots (Fig. 5). There are,
 192 however, significant differences between the invaded river plot and both the upland and cleared river
 193 plots for both *Prosopis* and *A. erioloba* (Tukey post hoc; $p < 0.001$).

194

195 **DISCUSSION**

196 Our belt transect results show that *Prosopis* density is highest in the riparian zone declining with
 197 distance from the river. The average groundwater depth in the river channel is 56 m, which is within the
 198 recorded range for *Prosopis* rooting (Canadell et al. 1996). While deep rooting may allow *Prosopis* access
 199 to deep water, *Prosopis* also has an extensive lateral root development which allows the genus to take
 200 advantage of sparse precipitation by growing outward and upward with roots extending to 5 cm of the soil
 201 surface (Gile et al. 1997). Combined with its extensive rooting system, the xylem anatomy of *Prosopis* is
 202 able to withstand a wide range in xylem pressure potentials which allow the species to grow in

203 environments where the water table may fluctuate over more than 4 m in a year (Stromberg 1992,
204 Pockman and Sperry 2000). This ability to utilise water at all depths in the soil combined with an ability
205 to adapt to all soil conditions makes *Prosopis* one of the most prolific invasives in low rainfall
206 environments (Gile et al. 1997).

207 Our belt transect results also show that the two native species *Ziziphus mucronata* and *Acacia*
208 *mellifera* are only evident in those transects where *Prosopis* had been cleared. We speculate that the
209 absence of these two species is because *Prosopis* is able to out-compete them for resources. Our stable
210 water isotope results show that in the riparian zone both *A. erioloba* and *Prosopis* are using the same
211 water source as both species use evaporatively enriched water during the wet season and deeper ground
212 water during the dry season when shallow water is depleted. This competition for water is evident in our
213 leaf $\delta^{13}\text{C}$ results which show less discrimination of the heavy ^{13}C isotope for *A. erioloba* in plots cleared
214 of *Prosopis*, indicating more water available to the plant. Xylem pressure potentials, an indicator of plant
215 moisture stress, show that both *Prosopis* and *A. erioloba* on the invaded riparian plot are more water
216 stressed than on both the cleared river plot and the upland plot.

217 There is a significant increase in dead material on *A. erioloba* in the invaded river plot relative to
218 the cleared river plot. We speculate that because *A. erioloba* is more water stressed in the invaded river
219 plot this increase in dead material is related to plant moisture stress. Our belt transect data show that
220 *Prosopis* abundance decreases significantly with distance from the river. In this upland area, where
221 distance to the water table is greater, *Prosopis* is not able to establish as easily as in the riparian zone.
222 With no significant differences in the amount of dead material and $\delta^{13}\text{C}$ values of *A. erioloba*, our results
223 show that this species is better adapted than *Prosopis* to accessing resources in the upland plots.. It is only
224 with an increase in competition for water with *Prosopis* (as in the riparian plots) that there is also a
225 significant increase in the amount of dead material on *A. erioloba*.

226 We believe that our results do show that there is competition for water between *Prosopis* and *A.*
227 *erioloba* at our study site. This competition for water results in *Prosopis* outcompeting *A. erioloba* for
228 water in the riparian zone resulting in severe canopy die back and even death of *A. erioloba*. Because of
229 its economic potential, there is reluctance among authorities to introduce biological control agents to
230 mitigate the spread of *Prosopis* in South Africa (Zachariades et al. 2011). Such reluctance can only be
231 overcome by studies such as ours that show the negative effects of *Prosopis* not only on available water
232 but also on species diversity and ecosystem functioning through the decline in viability of a keystone

233 species. Our study gives strong support for programs not only to remove *Prosopis* from rivers in the arid
 234 parts of South Africa but also for the release of well researched biological control agents (Zachariades et
 235 al. 2011).

236

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242

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322 **Figure Captions**

323 (Figures created in MS Office)

324

325 **Fig. 1** Results of the belt transect showing the decline in *Prosopis* density with distance from the river and
 326 the increase in both *Ziziphus mucronata* and *Acacia mellifera* in the cleared area. Values are for the total
 327 number of trees for three transects in and invaded and three transects in a cleared area

328

329 **Fig. 2** Mean and standard error for percentage canopy dieback showing the difference between upland
 330 and riparian *A. erioloba* trees where the amount of dead material increased in those trees in the invaded
 331 riparian plot but there were no significant differences between invaded upland and cleared upland plots

332

333 **Fig. 3** Mean $\delta^{18}\text{O}$ and $\delta^2\text{H}$ values (± 1 SE) for xylem water of *A. erioloba* for the dry season (\square), start of
 334 the wet season (\blacksquare) and end of the wet season (\blacksquare) for (a) cleared river, (b) cleared upland, (c) invaded river
 335 and (d) invaded upland. Also shown are values for xylem water of *Prosopis* for the dry season (\circ), start of
 336 the wet season (\bullet) and end of the wet season (\bullet). These values are plotted relative to the local meteoric
 337 water line (---) as well as average soil (\times), rain (\blacktriangle) and groundwater (\blacktriangle)

338

339 **Fig. 4** Mean and standard error for leaf $\delta^{13}\text{C}$ values for *A. erioloba* and *Prosopis* showing differences
 340 between upland and riparian trees

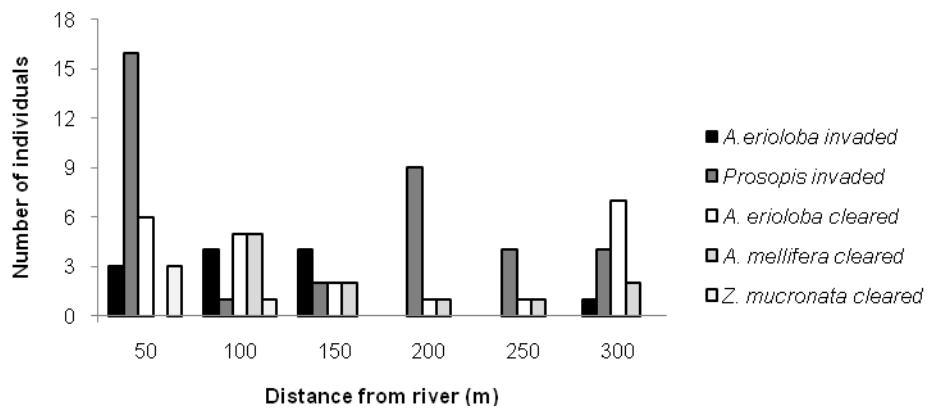
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342 **Fig. 5** Mean and standard error (n=6) predawn XPP's (MPa) for (a) cleared river (\square) and upland (\blacksquare) *A.*
 343 *erioloba*, (b) invaded river (\square) *A. erioloba* and invaded upland (\blacksquare) *A. erioloba* and (c) invaded river (\circ)
 344 *Prosopis* sp. and invaded upland (\bullet) *Prosopis* sp

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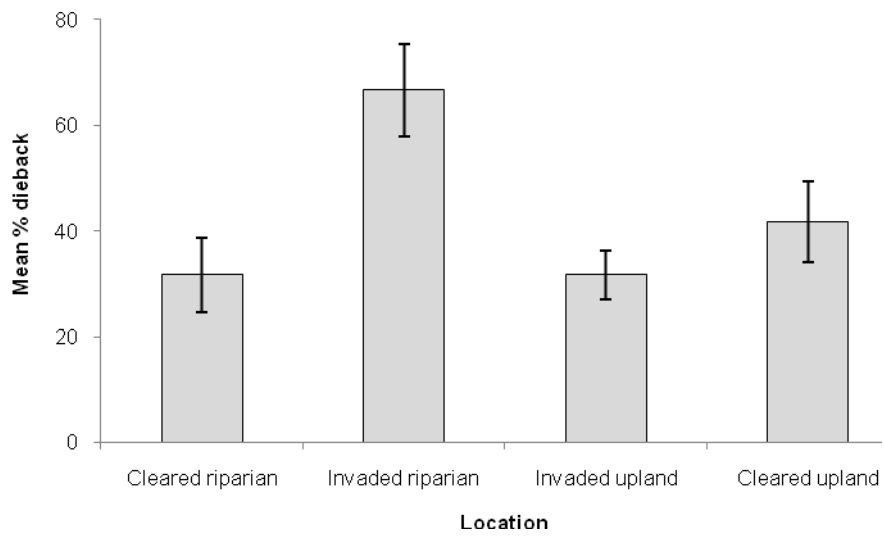
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351 **Fig. 1**

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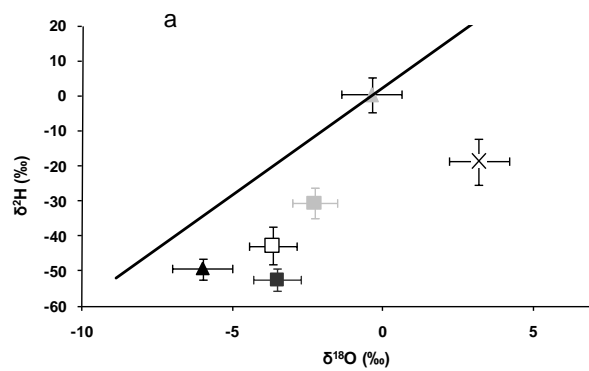


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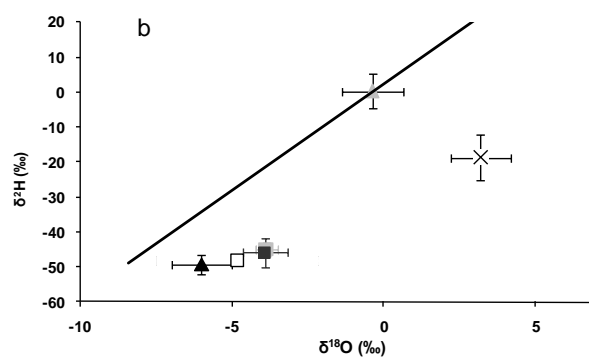
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356 **Fig. 2**

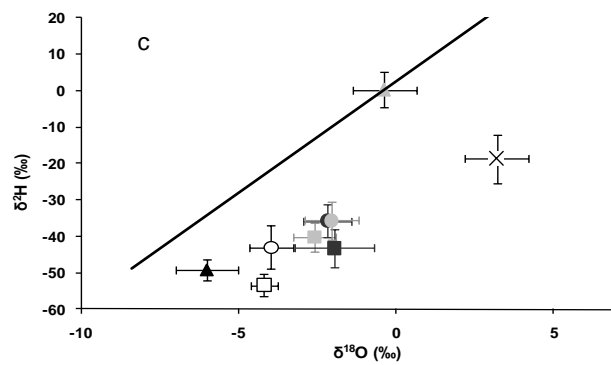
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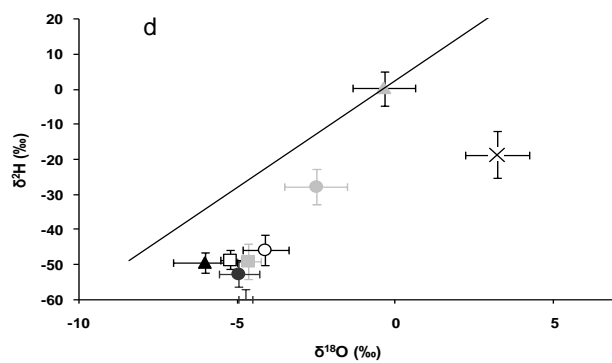
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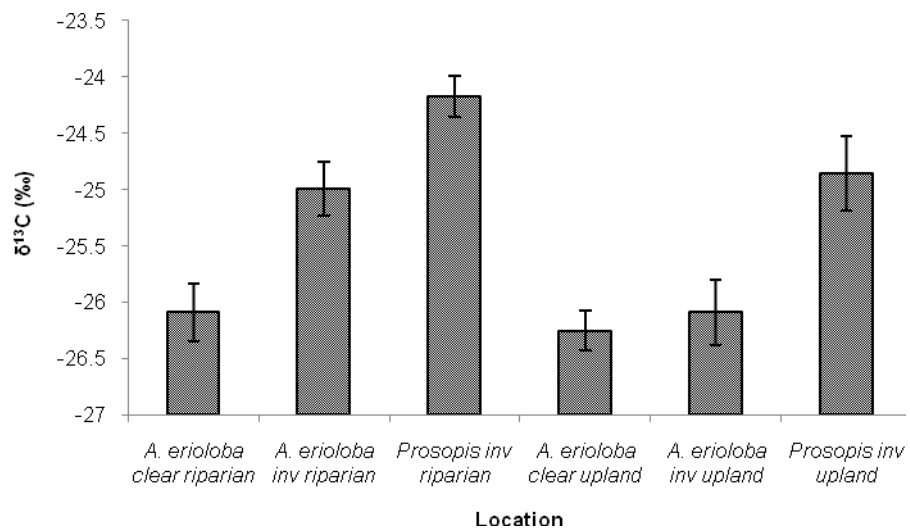


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363 **Fig. 3**

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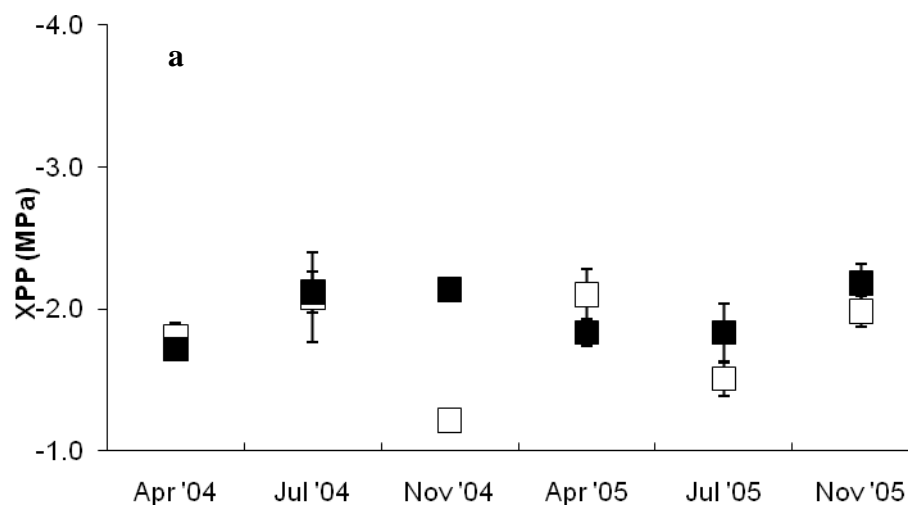


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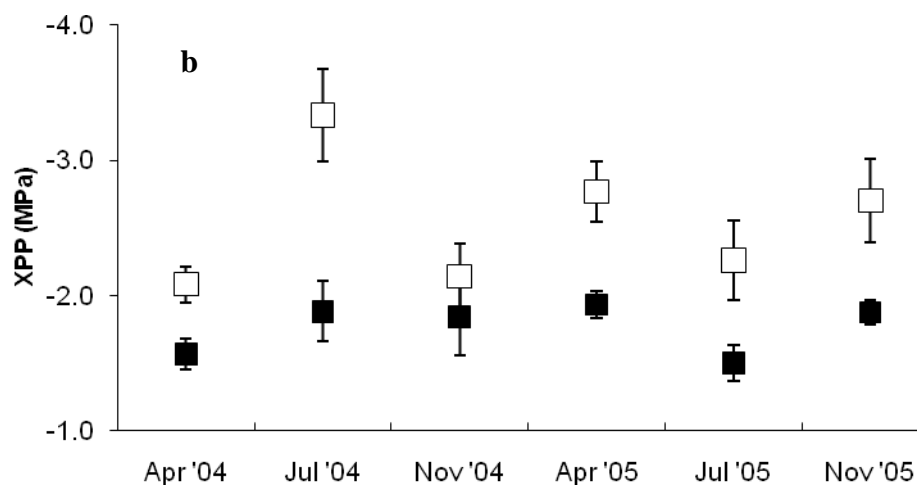
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367 **Fig. 4**

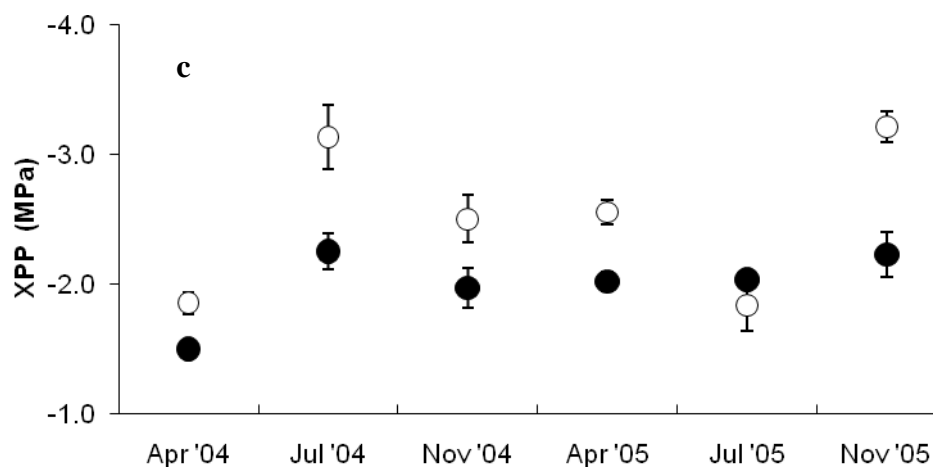
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373 **Fig. 5**

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